Kinetics of the Levels of Pro- and Anti-Inflammatory Cytokines and Vascular Endothelial Growth Factor in Serum and Pleural Fluid after Major Lung Resection for Lung Cancer

Yuji Taniguchi

Division of Organ Regeneration Surgery, Department of Surgery, School of Medicine, Tottori University, 683-8504 Japan

Some pro- and anti-inflammatory cytokines and angiogenesis-related factors play important roles in the inflammatory response after surgery. To study their kinetics and mutual relationships after major lung resection for lung cancer, concentrations of interleukin (IL)-6, -8 and -10 and vascular endothelial growth factor (VEGF) were measured in serum and pleural fluid. Venous blood and pleural fluid samples were collected before and up until 5 days after surgery from 10 patients with lung cancer, treated by standard lobectomy at Tottori University Hospital between 1997 and 1999. Cytokine levels after surgery were significantly higher than control levels before surgery in serum and pleural fluid. In pleural fluid, cytokines and VEGF increased after surgery 10 to 100 times than in serum. IL-6 and -10 reached their peaks in pleural fluid later than in serum, and then gradually decreased. Serum IL-8 reached its peak at 3 h, decreased until day 2 and then remained stable. Serum VEGF decreased until 3 h, and then showed little change. Pleural fluid IL-8 and VEGF reached their peaks at 3 or 6 h, decreased until day 2 and then increased up to day 5. The amount of cytokines produced per hour in drainage pleural fluid showed nearly the same kinetics as serum cytokines. Serum cytokines and VEGF showed higher coefficient values to the hourly production in pleural fluid than to the pleural fluid levels. During wound healing, IL-8 acted as a pro-inflammatory cytokine in the early phase, but as an angiogenic factor like VEGF in the late proliferative phase. The cytokine production per hour in pleural fluid is believed to be a useful marker of cytokine levels for evaluating surgical trauma after major lung resection for lung cancer.

Key words: cytokine; cytokine production per hour (hourly production of cytokine); lung cancer; vascular endothelial growth factor; wound healing

Interleukin (IL)-6 and IL-8 are known as proinflammatory cytokines, and IL-10 as an antiinflammatory cytokine. There have been many reports about the kinetics of pro- and antiinflammatory cytokine levels after surgery, including thoracic procedures (Steinberg et al., 1993; Sakamoto et al., 1994; Kawahito et al., 1995; Waller et al., 1996; Atwell et al., 1998; Liebold et al., 1999; Weissflog et al., 1999). However, most reports have described serum levels of cytokines, and few have reported their pleural fluid levels (Sakamoto et al., 1994; Weissflog et al., 1999). Thus, we thought it would be interesting to investigate the relationship of cytokine levels between serum and pleural fluid. In addition, vascular endothelial growth factor (VEGF) known as an important angiogenic factor (Fontanini et al., 1997; Inoue et al., 1997), has recently been demonstrated as acting together with pro- and anti-inflammatory cytokines (Ishimura et al., 1998; Nissen et al., 1998; Sato et al., 1999). In the present study, to clarify whe-

Abbreviations: IL, interleukin; VEGF, vascular endothelial growth factor

ther IL-6, IL-8, IL-10 and VEGF are associated after surgery for lung cancer, the kinetics of the levels of IL-6, IL-8, IL-10 and VEGF in both serum and pleural fluid were investigated.

Materials and Methods

Patients

The subjects were recruited from non-small cell lung cancer patients who underwent standard lobectomy by posterolateral thoracotomy with mediastinal nodal dissection in Tottori University Hospital between October 1997 and June 1999. We applied the following exclusion criteria for patient selection: i) incomplete resection, because the residual tumor has the potential to affect the cytokine levels (Matsuguchi et al., 1991; Yamamoto et al., 1996); ii) extended resection of the chest wall, diaphragm or mediastinum, because different degrees of surgical trauma have the potential to affect the cytokine levels (Sakamoto et al., 1994); iii) administration of perioperative ulinastatin or corticosteroid, because these agents influence cytokine release (Teoh et al., 1995; Kawamura et al., 1996) and iv) complication by pulmonary fibrosis, because this condition has the potential to affect the IL-8 level (Keane et al., 1997).

A total of 10 patients were examined in this study. They ranged in age from 49 to 76 years (mean, 64.3 years), consisting of 5 males and 5 females. Histologically, 3 patients suffered with squamous cell carcinomas, 6 patients with adenocarcinomas and 1 patient with mucoepidermoid carcinoma. Seven patients were in Stage I, one was in Stage II and two were in Stage III. All patients gave written informed consent. After tracheal intubation (double lumen endotracheal tube), general anesthesia was applied in combination with a thoracic epidural anesthesia in all patients. The thoracic epidural catheter was used for postoperative analgesia. Two thoracic tubes were indwelled in all patients, and the pleural fluid was constantly aspirated with -10 cmH₂O pressure. The duration of the surgery was 304.9 ± 13.3 min and the bleeding volume was 181.3 ± 57.3

mL. There were no severe postoperative complications such as respiratory failure, pneumonia, liver dysfunction or renal failure in these patients.

Blood and pleural fluid collection

Samples of venous blood were collected immediately after induction of anesthesia, and served as controls. Samples of pleural fluid were collected by intrathoracic lavage with 50 mL of physiological saline just after thoracotomy, and used to determine the preoperative control data of pleural fluid.

The other samples of serum and pleural fluid were simultaneously collected at 3 and 6 h and on days 1, 2, 3 and 5 after surgery. On days 1, 2, 3 and 5, the samples were collected 9:00 in the morning. All samples were centrifuged immediately at 2,500 rpm for 15 min at 4°C. Centrifuged samples were stored at -30° C until the assays were performed.

Cytokine assays

IL-6, IL-8, IL-10 and VEGF concentrations were measured using commercial enzymelinked immunosorbent assay kits (Genzyme, Cambridge, MA for IL-6 and IL-10; Endogen, Woburn, MA for IL-8; Techne, Minneapolis, MN for VEGF).

Statistical analysis

Statistical analysis was performed with Stat-View 5.0 J software (Abacus, Concepts, Inc., Berkeley, CA). Differences in the kinetics of cytokine and VEGF levels were analyzed using Friedman's test and Wilcoxon's signed rank test. Correlations between the serum and pleural fluid levels of cytokines and VEGF were analyzed using Spearman's test. Results are expressed as mean \pm SEM. *P* values of < 0.05 were considered significant.



Results

Kinetics of serum cytokine and VEGF levels

Serum levels of cytokines after surgery (Figs. 1A–C) were significantly higher than control levels throughout. IL-6 (Fig. 1A) and IL-10 (Fig. 1C) reached their peaks at 3 h (IL-6: $626.8 \pm 63.0 \text{ pg/mL}$, IL-10: $15.9 \pm 1.6 \text{ pg/mL}$) and gradually declined with time. Serum IL-8 (Fig. 1B) showed its peak at 3 h (47.5 ± 12.0 pg/mL), declined until day 2 and then remained stable. However, serum VEGF levels (Fig. 1D) decreased until 3 h, followed by little change.

Figs. 1A–D. Kinetics of serum levels of cytokines and VEGF measured before and after lung resection. *P < 0.05 and **P < 0.01compared with the preoperative control level. The data are presented as mean ± SEM. IL, interleukin; VEGF, vascular endothelial growth factor.

Y. Taniguchi



Kinetics of drainage fluid cytokine and VEGF levels

Pleural fluid levels of cytokines and VEGF (Figs. 2A-D) after surgery were significantly higher than control levels throughout. IL-6 and IL-8 were about 100-fold and IL-10 and VEGF were about 10-fold higher in concentration than in serum. Concerning level changes, IL-6 (Fig. 2A) and IL-10 (Fig. 2C) reached their peaks (IL-6: 177,000 ± 30,600 pg/mL, IL-10: 481.0 ± 91.8 pg/mL) 6 h or 1 day after surgery, later than in the patterns seen in serum. In pleural fluid, however, IL-8 (Fig. 2B) and VEGF (Fig. 2D) levels showed their peaks in the early phases, at 3 or 6 h (IL-8: 19,700 ± 5030 pg/mL, VEGF: 1838 ±

Figs. 2A–D. Kinetics of pleural fluid levels of cytokines and VEGF measured before and after lung resection. *P < 0.05 and **P < 0.01 compared with the preoperative control level. The data are presented as mean ± SEM. IL, interleukin; VEGF, vascular endothelial growth factor.



182 pg/mL), decreased until day 2 (IL-8: 2286 \pm 787 pg/mL, VEGF: 980 \pm 223 pg/mL) and elevated again during the late phase up to day 5 (IL-8: 14,900 \pm 9580 pg/mL, VEGF: 1553 \pm 261 pg/ mL). When the amount of cytokines produced per hour (hourly production) in pleural fluid was calculated by multiplying the concentration by drainage volume per hour, the hourly cytokine production in pleural fluid (Figs. 3A– C) showed nearly the same kinetics as serum concentration kinetics (Figs. 1A–C).

Figs. 3A–D. Kinetics of hourly production in pleural fluid of cytokines and VEGF measured before and after lung resection. *P < 0.05 and **P < 0.01 compared with the preoperative control level. The data are presented as mean ± SEM. IL, interleukin; VEGF, vascular endothelial growth factor.



Relationships of cytokine and VEGF levels between serum and pleural fluid

All factors showed significant correlations between serum levels and hourly productions in pleural fluid (IL-6: r = 0.777, IL-8: r = 0.550, IL-10: r = 0.580, VEGF: r = 0.264) (Figs. 4A–D). These values of coefficients were higher than those between serum levels and pleural fluid levels for all factors (IL-6: r = 0.517, IL-8: r = 0.494, IL-10: r = 0.564, VEGF: r = 0.235).

Discussion

Various cytokines with a wide range of biological activities play the role of controlling inflammatory and immunological reactions to surgical trauma. In the present study, the concentrations of cytokines in pleural fluid were about 100-fold greater than those in serum. Previous researchers also reported that the IL-6 and IL-8 levels in thoracic drainage fluid were about 100- to 1000-fold greater than those in peripheral blood (Sakamoto et al., 1994; Krohn et al., 1998). Therefore, it is very clear that the cytokine response is much stronger in the local pleural cavity than in the systemic blood.

Figs. 4A–D. Relationships between serum levels and hourly productions in pleural fluid studied for cytokines and VEGF. IL, interleukin; VEGF, vascular endothelial growth factor.

In our present study, the kinetics of cytokine levels in serum and pleural fluid showed significant correlations. In particular, the correlation between the hourly cytokine production in pleural fluid and the serum cytokine level was higher than that between the pleural fluid cytokine level and the serum cytokine level. Sakamoto et al. (1994) reported that the messenger RNAs of cytokines could not be detected in leukocytes from the peripheral blood but could be demonstrated in leukocytes from drainage fluid: they suggested that cytokines are induced and secreted mainly in the surgical field and simultaneously appear in the blood stream. Asadullah et al. (1995) reported data from neurosurgery supporting this hypothesis. Regarding the mechanism of this effect, we consider it possible that pleural cytokines spill over into the peripheral blood through pleural absorption. Therefore, when the concentrations of cytokines in pleural fluid are markedly higher than serum cytokine levels, such as after thoracic surgery, serum cytokine levels are strongly influenced by the amount of cytokines produced per hour in pleural fluid. Pleural fluid can be collected because the thoracic tube is always indwelled after thoracic surgery. Thus, the hourly production of cytokines in pleural fluid appears to be a new useful marker of cytokine levels for evaluating the degree of trauma caused by thoracic procedures.

IL-8 is a pro-inflammatory cytokine and has a potent chemoattractant activity for neutrophils (Baggiolini et al., 1989). Recent studies have shown that the serum IL-8 level peaked within 24 h after surgery and decreased thereafter (Sakamoto et al., 1994; Atwell et al., 1998; Liebold et al., 1999). However, few studies have examined the kinetics of IL-8 later than 3 days after surgery, particularly after major lung surgery. In the present study, the kinetics of the IL-8 level in pleural fluid showed 2 significant peaks postoperatively, at 3 or 6 h and again on day 5. Until day 2 after surgery, IL-8 functioned as a pro-inflammatory cytokine. However, re-elevation of IL-8 later than day 3 did not necessarily indicate its function as a proinflammatory cytokine at that time. Koch et al. (1992) reported that IL-8 has another function; it induces neovascularization without inflammation. During wound healing, the period until day 3 after surgery corresponds to an inflammatory phase, while the period later than day 3 corresponds to a proliferative phase (Fine and Mustoe, 1997) during which angiogenesis occurs at the wound site (Adzick, 1997). Grad et al. (1998) reported that, in multiple trauma patients, serum VEGF was elevated from day 3 after injury. Angiogenesis-related factors such as VEGF are induced in hypoxia (Shweiki et al., 1992; Steinbrech et al., 1999). Hangai (1996) reported that the tissue oxygen tension at the anastomosis site reached its lowest level on day 3 after tracheoplasty. Therefore, this condition may re-elevate VEGF levels after day 3. Because both IL-8 and VEGF showed the same kinetics in pleural fluid, we figured that the function of IL-8 later than day 3 after surgery could act as an angiogenic factor rather than as a proinflammatory cytokine.

In the present study, IL-8 and VEGF were produced at high levels in pleural fluid after lung surgery. Angiogenesis is required for the growth and metastasis of solid tumors (Folkman et al., 1989; Bicknell and Harris, 1991; Imoto et al., 1998), and anti-IL-8 and anti-VEGF therapy can inhibit both primary tumor growth and metastasis (Asano et al., 1995; Warren et al., 1995; Arenberg et al., 1996; Rowe et al., 2000). Moreover, the inflammatory cytokine level in serum was reported to be higher after lobectomy of the lung than by gastrectomy and colorectal resection (Sakamoto et al., 1994). Proinflammatory cytokines such as IL-6 and IL-8 stimulate VEGF expression (Hanahan and Folkman, 1996; Risau, 1997). It has been reported that excess surgical stress from thoracotomy facilitates metastasis (Hattori et al., 1980; Hirai et al., 1997). Furthermore, Gabrilovich et al. (1996) recently reported that in vivo VEGF can inhibit the functional maturation of dendritic cells which play a critical role in antitumor immune responses. Based on the above, lobectomy by standard thoracotomy for lung cancer may likely induce proliferation and metastasis of the tumor not only due to angiogenesis but also to tumor immunity because VEGF was produced at high levels after lung resection.

Our study examined only a small number of patients who underwent elective surgery for lung cancer. Also, because none of our patients had severe postoperative complications, our findings only reflected normal cytokine kinetics after major lung resection for lung cancer. Therefore, further studies are needed to investigate whether our findings are applicable to wedge resection, to lobectomy under videoassisted thoracic surgery regarded as minimally invasive for lung cancer, to extended procedures such as pneumonectomy and combined resection of other involved organs for primary lung cancer, and to lung surgery for conditions other than malignancy. If the cytokine cascade producing the pro- and anti-inflammatory and angiogenic reactions were balanced, normal healing after surgical trauma could be achieved. However, overexpression of angiogenic factors such as IL-8 and VEGF might promote recurrence or metastasis of cancer cells. Therefore, studies of cytokine kinetics will facilitate understanding of the normal biological response and provide reference information for optimal perioperative patient management.

Acknowledgments: The author would like to thank Professor Yasuaki Kawai of the Division of Adaptation Physiology, Department of Functional, Morphological and Regulation Science, Professor Shigetsugu Ohgi of the Division of Organ Regeneration Surgery, Department of Surgery and Professor Eiji Shimizu of the Division of Internal Medicine and Molecular Therapeutics, Department of Multidisciplinary Internal Medicine, School of Medicine, Tottori University for their kind advice and valuable suggestions concerning this study.

References

- Adzick NS. Wound healing: biologic and clinical features. In: Sabiston DC Jr, ed. Textbook of surgery: the biological basis of modern surgical practice. 15th ed. Philadelphia: WB Saunders Co.; 1997. p. 207–220.
- 2 Arenberg DA, Kunkel SL, Polverini PJ, Glass M, Burdick MD, Strieter RM. Inhibition of interleukin-8 reduces tumorigenesis of human non-small cell lung cancer in SCID mice. J Clin Invest 1996;97: 2792–2802.
- 3 Asadullah K, Woiciechowsky C, Döcke WD,

Liebenthal C, Wauer H, Kox W, et al. Immunodepression following neurosurgical procedures. Crit Care Med 1995;23:1976-1983.

- 4 Asano M, Yukita A, Matsumoto T, Kondo S, Suzuki H. Inhibition of tumor growth and metastasis by an immunoneutralizing monoclonal antibody to human vascular endothelial growth factor/vascular permeability factor₁₂₁. Cancer Res 1995;55:5296–5301.
- 5 Atwell DM, Grichnik KP, Newman MF, Reves JG, McBride WT. Balance of proinflammatory and antiinflammatory cytokines at thoracic cancer operation. Ann Thorac Surg 1998;66:1145–1150.
- 6 Baggiolini M, Walz A, Kunkel S. Neutrophilactivating peptide-1/interleukin 8, a novel cytokine that activates neutrophils. J Clin Invest 1989;84: 1045–1049.
- 7 Bicknell R, Harris AL. Novel growth regulatory factors and tumour angiogenesis. Eur J Cancer 1991;27:781–785.
- 8 Fine NA, Mustoe TA. Wound healing. In: Greenfield LJ, ed. Surgery: scientific principles and practice. 2nd ed. Philadelphia: Lippincott-Raven; 1997. p. 67–83.
- 9 Folkman J, Watson K, Ingber D, Hanahan D. Induction of angiogenesis during the transition from hyperplasia to neoplasia. Nature 1989;339: 58–61.
- 10 Fontanini G, Vignati S, Boldrini L, Chiné S, Silvestri V, Lucchi M, et al. Vascular endothelial growth factor is associated with neovascularization and influences progression of non-small cell lung carcinoma. Clin Cancer Res 1997;3:861– 865.
- 11 Gabrilovich DI, Chen HL, Girgis KR, Cunningham HT, Meny GM, Nadaf S, et al. Production of vascular endothelial growth factor by human tumors inhibits the functional maturation of dendritic cells. Nature Med 1996;2:1096–1103.
- 12 Grad S, Ertel W, Keel M, Infanger M, Vonderschmitt DJ, Maly FE. Strongly enhanced serum levels of vascular endothelial growth factor (VEGF) after polytrauma and burn. Clin Chem Lab Med 1998; 36:379–383.
- 13 Hanahan D, Folkman J. Patterns and emerging mechanisms of the angiogenic switch during tumorigenesis. Cell 1996;86:353–364.
- 14 Hangai N. Change of tissue oxygen tension and blood flow of the trachea after tracheoplasty. Kikanshigaku 1996;18:646–653 (in Japanese with English abstract).
- 15 Hattori T, Hamai Y, Takiyama W, Hirai T, Ikeda T. Enhancing effect of thoracotomy on tumor growth in rats with special reference to the duration and timing of the operation. Gann (Jpn J Cancer Res) 1980;71:280–284.
- 16 Hirai T, Yoshimoto A, Iwata T, Yamashita Y, Kuwahara M, Toge T. Enhancing effect of thoraco-laparotomy on liver metastasis and the

role played by active oxygens in its mechanism. Surg Today (Jpn J Surg) 1997;27:1040–1045.

- 17 Imoto H, Osaki T, Taga S, Ohgami A, Ichiyoshi Y, Yasumoto K. Vascular endothelial growth factor expression in non-small-cell lung cancer: prognostic significance in squamous cell carcinoma. J Thorac Cardiovasc Surg 1998;115:1007–1014.
- 18 Inoue K, Ozeki Y, Suganuma T, Sugiura Y, Tanaka S. Vascular endothelial growth factor expression in primary esophageal squamous cell carcinoma: association with angiogenesis and tumor progression. Cancer 1997;79:206–213.
- 19 Ishimura K, Tsubouchi T, Okano K, Maeba T, Maeta H. Wound healing of intestinal anastomosis after digestive surgery under septic conditions: participation of local interleukin-6 expression. World J Surg 1998;22:1069–1076.
- 20 Kawahito K, Kawakami M, Fujiwara T, Adachi H, Ino T. Interleukin-8 and monocyte chemotactic activating factor responses to cardiopulmonary bypass. J Thorac Cardiovasc Surg 1995; 110:99–102.
- 21 Kawamura T, Inada K, Akasaka N, Wakusawa R. Ulinastatin reduces elevation of cytokines and soluble adhesion molecules during cardiac surgery. Can J Anaesth 1996;43:456–460.
- 22 Keane MP, Arenberg DA, Lynch JP III, Whyte RI, Iannettoni MD, Burdick MD, et al. The CXC chemokines, IL-8 and IP-10, regulate angiogenic activity in idiopathic pulmonary fibrosis. J Immunol 1997;159:1437–1443.
- 23 Koch AE, Polverini PJ, Kunkel SL, Harlow LA, DiPietro LA, Elner VM, et al. Interleukin-8 as a macrophage-derived mediator of angiogenesis. Science 1992;258:1798–1801.
- 24 Krohn CD, Reikerås O, Mollnes TE, Aasen AO. Complement activation and release of interleukin-6 and tumour necrosis factor-α in drained and systemic blood after major orthopaedic surgery. Eur J Surg 1998;164:103–108.
- 25 Liebold A, Keyl C, Birnbaum DE. The heart products but the lungs consume proinflammatory cytokines following cardiopulimonary bypass. Eur J Cardiothorac Surg 1999;15:340–345.
- 26 Matsuguchi T, Okamura S, Kawasaki C, Shimoda K, Omori F, Hayashi S, et al. Constitutive production of granulocyte colony-stimulating factor and interleukin-6 by a human lung cancer cell line, KNSY: gene amplification and increased mRNA stability. Eur J Haematol 1991;47:128–133.
- 27 Nissen NN, Polverini PJ, Koch AE, Volin MV, Gamelli RL, DiPietro LA. Vascular endothelial growth factor mediates angoigenic activity during the proliferative phase of wound healing. Am

J Pathol 1998;152:1445-1452.

- 28 Risau W. Mechanisms of angiogenesis. Nature 1997;386:671–674.
- 29 Rowe DH, Huang J, Kayton ML, Thompson R, Troxel A, O'Toole KM, et al. Anti-VEGF antibody suppresses primary tumor growth and metastasis in an experimental model of Wilms' tumor. J Pediatr Surg 2000;35:30–33.
- 30 Sakamoto K, Arakawa H, Mita S, Ishiko T, Ikei S, Egami H, et al. Elevation of circulating interleukin 6 after surgery: factors influencing the serum level. Cytokine 1994;6:181–186.
- 31 Sato Y, Oshima T, Kondo Y. Regulatory role of endogenous interleukin-10 in cutaneous inflammatory response of murine wound healing. Biochem Biophys Res Commun 1999;265:194–199.
- 32 Shweiki D, Itin A, Soffer D, Keshet E. Vascular endothelial growth factor induced by hypoxia may mediate hypoxia-initiated angiogenesis. Nature 1992;359:843–845.
- 33 Steinberg JB, Kapelanski DP, Olson JD, Weiler JM. Cytokine and complement levels in patients undergoing cardiopulmonary bypass. J Thorac Cardiovasc Surg 1993;106:1008–1016.
- 34 Steinbrech DS, Mehrara BJ, Chau D, Rowe NM, Chin G, Lee T, et al. Hypoxia upregulates VEGF production in keloid fibroblasts. Ann Plast Surg 1999;42:514–520.
- 35 Teoh KHT, Bradley CA, Gauldie J, Burrows H. Steroid inhibition of cytokine-mediated vasodilation after warm heart surgery. Circulation 1995; 92:347–353.
- 36 Waller DA, Keavey P, Woodfine L, Dark JH. Pulmonary endothelial permeability changes after major lung resection. Ann Thorac Surg 1996;61:1435–1440.
- 37 Warren RS, Yuan H, Matli MR, Gillett NA, Ferrara N. Regulation by vascular endothelial growth factor of human colon cancer tumorigenesis in a mouse model of experimental liver metastasis. J Clin Invest 1995;95:1789–1797.
- 38 Weissflog D, Kroegel C, Luttmann W, Grahmann PR, Hasse J. Leukocyte infiltration and secretion of cytokines in pleural drainage fluid after thoracic surgery. Chest 1999;115:1604–1610.
- 39 Yamamoto Y, Toi M, Kondo S, Matsumoto T, Suzuki H, Kitamura M, et al. Concentrations of vascular endothelial growth factor in the sera of normal controls and cancer patients. Clin Cancer Res 1996;2:821–826.

Received April 19, 2002; accepted May 13, 2002

Corresponding author: Dr. Yuji Taniguchi