articular cartilage were positive for IGF-1 and IGF-1 receptor in all animals with osteoarthritis in each grade. In the moderate group, in which articular cartilage degeneration had progressed further, the presence of IGF-1 and IGF-1 receptor was noted not only in fibroblastic-like cells, but also in chondrocytes clustering in a layer deeper than fibroblastic-like cells. In contrast to chondrocytes in the weight-bearing regions, these chondrocytes proved to have a very high mitotic activity in this study. Therefore, it can be speculated that chondrocytes presumed to be differentiated from totipotent immature mesenchymal cells may have produced a cartilage matrix by the autocrine mechanism via IGF-1. A previous report has demonstrated that more IGF-1 is present in human osteoarthritis cartilage and synovial fluid than in normal cartilage and synovial fluid, and suggested that this IGF-1 originated from the synovial membrane (Schneiderman et al., 1995). However, this study could not demonstrate that IGF-1 secreted by totipotent immature mesenchymal cells directly participated in the differentiation to chondrocytes. Some studies have indicated the close involvement of TGF-beta 1 in experimental osteophyte formation (van Beuningen et al., 1994; van den Berg, 1995), and speculated that TGFbeta 1 has a facilitatory effect on differentiation of totipotent immature mesenchymal cells. Furthermore, other growth factors are also thought to interact in a complicated manner (Trippel, 1995).

Acknowledgments: We would like to thank Prof. Ryota Teshima, Dept. of Orthopedic Surgery, Faculty of Medicine, Tottori Univ., for advice and supervision. I would also like to thank other members of the Department for their advice and help.



Fig. 4. Safranin-O (**A**) staining discloses the obvious formation of an osteophyte protruding from the articular margin in the severe group of 18-month-old animals with end-stage osteoarthritis. The osteophyte consists of type-I collagen (**B**), and hardly any chondrocytes and cartilage matrix positive for type-II collagen are observed (**C**). Fibrous connective tissue (arrows) is noted around the osteophyte, and IGF-1 (**F**) and IGF-1 receptor (**G**) are identified in the fibrous connective tissue. Fibroblastic-like cells in the fibrous connective tissue are positive for both type-I (**B**) and -III (**D**) collagens, and slightly positive for PCNA (**E**). JS, joint space; T, tibia. Original magnification: A, × 100; B–G, × 200.

[Figs. 4A-C on p. 138; Figs. 4D-G on p. 139]



Figs. 4D–G. Continued from the previous page.

References

- Aigner T, Bertling W, Stöß H, Weseloh G, von der Mark K. Independent expression of fibril-forming collagens I, II, and III in chondrocytes of human osteoarthritic cartilage. J Clin Invest 1993;91:829–837.
- 2 Aigner T, Dietz U, Stöß H, von der Mark K. Differential expression of collagen types I, II, III, and X in human osteophytes. Lab Invest 1995;73:236–243.
- 3 Allard SA, Bayliss MT, Maini RN. The synovium-cartilage junction of the normal human knee. Implications for joint destruction and repair. Arthritis Rheum 1990;33: 1170–1179.
- 4 Ash P, Francis MJO. Response of isolated rabbit articular and epiphyseal chondrocytes to rat liver somatomedin. J Endocrinol 1975;66:71–78.
- 5 Bendele AM, White SL, Hulman JF. Osteoarthrosis in guinea pigs: histopathologic and scanning electron microscopic features. Lab Anim Sci 1989;39:115–121.
- 6 Bluestone R, Bywaters EGL, Hartog M, Holt PJL, Hyde S. Acromegalic arthropathy. Ann Rheum Dis 1971;30:243–258.
- 7 de Bri E, Jönsson K, Reinholt FP, Svensson O. Focal destruction and remodeling in guinea pig arthorosis. Acta Orthop Scand 1996;67:498–504.
- 8 Jaffe HL. Metabolic, Degenerative and inflammatory diseases of bones and joints. Philadelphia: Lea & Febiger; 1972.
- 9 Jewell FM, Watt I, Doherty M. Plain radiographic features of osteoarthritis. In: Brandt KD, Doherty M, Lohmander LS, eds. Osteoarthritis. New York: Oxford University Press; 1998. p. 217–237.
- 10 Johanson NA, Vigorita VJ, Goldman AB, Salvati EA. Acromegalic arthropathy of the hip. Clin Orthop 1983;173:130–139.
- 11 LeRoith D, Kavsan VM, Koval AP, Roberts CT Jr. Phylogeny of the insulin-like growth factors (IGFs) and receptors: a molecular approach. Mol Reprod Dev 1993;35:332– 338.
- 12 Lieberman SA, Björkengren AG, Hoffman AR. Rheumatologic and skeletal changes in acromegaly. Endocrinol Metab Clin North Am 1992;21:615–631.
- 13 Linsenmayer TF, Toole BP, Trelstad RL. Temporal and spatial transitions in collagen types during embryonic chick limb development. Dev Biol 1973;35:232–239.
- 14 Luyten FP, Hascall CV, Nissley SP, Morales TI, Reddi AH. Insulin-like growth factors

maintained steady state metabolism of proteoglycans in bovine articular cartilage explants. Arch Biochem Biophys 1988;267:416–425.

- 15 Mankin HJ, Dorfman H, Lippiello L, Zarins A. Biochemical and metabolic abnormalities in articular cartilage from osteo-arthritic human hips. J Bone Joint Surg 1971;53-A:523–537.
- 16 McQuillan DJ, Handley CJ, Campbell MA, Bolis S, Milway VE, Herington AC. Stimulation of proteoglycan biosynthesis by serum and insulinlike growth factor-I in cultured bovine articular cartilage. Biochem J 1986;240:423–430.
- 17 Moskowitz RW, Goldbeg VM. Studies of osteophyte pathogenesis in experimentally induced osteoarthritis. J Rheumatol 1987;14:311–320.
- 18 Okada Y, Shinmei M, Tanaka O, Naka K, Kimura A, Nakanishi I et al. Localization of matrix metalloproteinase 3 (stromelysin) in osteoarthritic cartilage and synovium. Lab Invest 1992; 66:680–690.
- 19 Resnick D. Pituitary disorders. In: Resnick D, Niwayama G, eds. Diagnosis of bone and joint disorders. Philadelphia: W. B. Saunders Company; 1988. p. 2172–2198.
- 20 Schneiderman R, Rosenberg N, Hiss J, Lee P, Liu F, Hintz RL et al. Concentration and size distribution of insulin-like growth factor-I in human normal and osteoarthritic synovial fluid and cartilage. Arch Biochem Biophys 1995;324:173–188.
- 21 Telhag H, Lindberg L. A method for inducing osteoarthritic changes in rabbits' knees. Clin Orthop 1972;86:214–223.

- 22 Tokuda M. Histological study of spontaneous osteoarthritis in the knee joint of guinea pigs. J Orthop Sci 1997;2:248–258.
- 23 Trippel SB, Corvol MT, Dumontier MF, Rappaport R, Hung HH, Mankin HJ. Effect of somatomedin-C/insulin-like growth factor I and growth hormone on cultured growth plate and articular chondrocytes. Pediatr Res 1989;25:76–82.
- 24 Trippel SB. Growth factor action on articular cartilage. J Rheumatol 1995;22:129–132.
- 25 van Beuningen HM, van der Kraan PM, Arntz OJ, van den Berg WB. Transforming growth factor-beta 1 stimulates articular chondrocyte proteoglycan synthesis and induces osteophyte formation in the murine knee joint. Lab Invest 1994;71:279–290.
- 26 van den Berg WB. Growth factors in experimental osteoarthritis: Transforming growth factor beta pathogenic? J Rheumatol 1995;22: 143–145.
- 27 von der Mark K, von der Mark H, Gay S. Study of differential collagen synthesis during development of the chick embryo by immunofluorescence. Dev Biol 1976;53:153–170.
- 28 von der Mark K, von der Mark H. The role of three genetically distinct collagen types in endochondral ossification and calcification of cartilage. J Bone Joint Surg 1977;59-B:458–464.

Received September 4, 2000; accepted October 24, 2000

Corresponding author: Dr. Masanori Ono